

● **Core Facility for High Throughput Cell Based Screens**
 Biotech Research and Innovation Center (BRIC), Faculty of Health and Medical Sciences
 Copenhagen University

<https://www.bric.ku.dk/core-facilities/ht-cell-based-screens/>

DENMARK



specialist
screening site

COPENHAGEN

a nominated partner site of



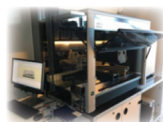
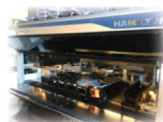
THE PEOPLE

Krister Wennerberg, Prof
Cornelia Steinhauer, PhD
Elin Pietras, PhD
Henning Gram Hansen, PhD

THE HARDWARE

HTS screening systems

Automated liquid handling is primarily performed on a STAR and STARlet liquid handling station (Hamilton) equipped with a 96well and 384well head respectively and plate handling capacities. Low volumes and more flexible liquid transfer are performed on an Echo 550 acoustic dispenser (Labcyte). In addition, the facility has access to an Echo525 (Labcyte), MultiFlo FX dispenser (Biotek), ELX 405 washer (Biotek) and Mantis Microfluidic Liquid Handler (Formulatrix).



Instrument
Manufacturer
Capabilities

Microlab STAR
Hamilton
Liquid transfer using disposable pipettips (Hamilton); 96well head and individual channels; vacuum manifold; shaker; plate handling; barcode reader

Microlab STARlet
Hamilton
Liquid transfer using disposable pipettips (Hamilton); 384well head; plate handling

Echo 550
Labcyte
Liquid transfer using acoustic dispensing; 2.5nl
any-well-to-any-well transfer

Applications

HT 96well
transfections/treatments;
miniprep; library replating
96- and 384-well

HT 384well
transfections/treatments; HT-
QPCR; library handling
384-well

HT 384well compound transfer;
library cherry pick

Plate format

96- and 384-well

384-well

2.5nl-2µl

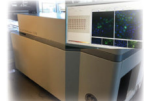
Transfer volume

2-1000µl

2-50µl

2.5nl-2µl

Readouts/ Screening technologies
 Phenotypic cell-based readouts are primarily performed on an IN Cell Analyzer 2200 high content microscope (GE Healthcare). Life imaging can be followed on either our ZOOM or S3 Incucyte systems (Sartorius). For absorbance or luminescence based assays we use a Synergy HTX Multi-Mode Reader (Biotek). In addition, the facility has access to an ENSIGHT plate reader with imaging capacity (PerkinElmer) and an iQue Screener Plus flow cytometer (Sartorius) for suspension cells.



Instrument
Manufacturer
Capabilities

IN Cell Analyzer 2200
GE Healthcare Life Sciences
Fast, sensitive, and flexible
automated cell imaging using
4x, 10x, 20x and 40x objectives
and filters for imaging DAPI,
FITC, Cy3, TxRed, Cy5, CFP and
YFP; non-confocal but with
advanced 3D deconvolution
software

Synergy HTX Multi-Mode Reader
BioTek
Flexible and sensitive plate reader with
monochromator based absorbance and filter
based fluorescence/luminescence

Applications

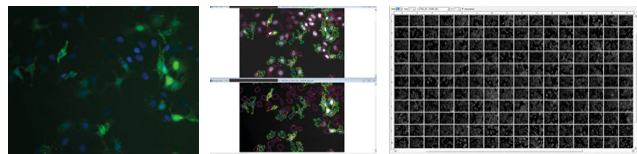
Broad range of high-content
assays, 2D and 3D
slide to 384-well

Broad range of absorbance or luminescence based
biological readouts; DNA measurement
96- and 384-well

THE SOFTWARE

Data analysis tools

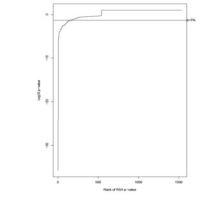
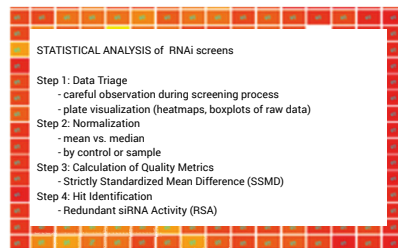
The Core Facility for HT Cell Based Screens at BRIC offers cell-based assay development and high content screening services utilizing phenotypic image-based analysis as primary readout. The facilities IN Cell Analyzer 2200 automated high content microscope (GE Healthcare) is a non-confocal system with advanced 3D deconvolution software. It allows for fast, sensitive, and flexible automated cell imaging using 4x, 10x, 20x and 40x objectives and filters for imaging DAPI, FITC, Cy3, TxRed, Cy5, CFP and YFP. The proprietary IN Cell Investigator and INCarta software provides sophisticated image analysis on various user levels. In addition, we utilize Velocity (PerkinElmer) and Cell Profiler Analyst (Broad Institute) for advanced image analysis.



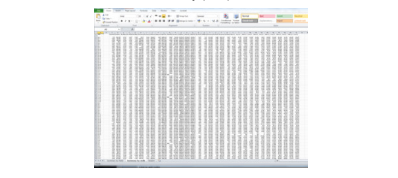
Images courtesy of Roland Baumgartner and Alison Low

Software tools

The facility has developed their own R-based statistical pipeline and data analysis workflow utilizing RSA analysis as hit picking algorithm (2). Data visualization and further statistical analyses are performed using Spotfire High Content Profiler (PerkinElmer) and JMP (SAS).



Birmingham et al. Nat Methods 2009; 6(8): 569-575



THE OUTPUT

1. eIF5A is required for autophagy by mediating ATG3 translation. Lubas M, Harder LM, Kumsta C, Tiesens I, Hansen M, Andersen JS, Lund AH, Frankel LB. EMBO Rep. 2018 Jun;19(6). pii: e46072. doi: 10.15252/embr.201846072. Epub 2018 Apr 30.
2. High-throughput siRNA screening applied to the ubiquitin-proteasome system. Poulsen EG, Nielsen SV, Pietras EJ, Johansen JV, Steinhauer C and Hartmann-Petersen R Methods in Molecular Biology. Proteostasis 2016; 1449:421-39
3. miR-339-5p regulates the p53 tumor-suppressor pathway by targeting MDM2. Jansson MQ, Damas ND, Lees M, Jacobsen A, Lund AH. Oncogene. 2014 Jun 2. doi: 10.1038/nc.2014.130.
4. A Screen Identifies the Oncogenic Micro-RNA miR-378a-5p as a Negative Regulator of Oncogene-Induced Senescence. Kooistra SM, Nørgaard LC, Lees MJ, Steinhauer C, Johansen JV, Helin K. PLoS One. 2014 Mar 20;9(3):e91034. doi: 10.1371/journal.pone.0091034. eCollection 2014
5. Histone acetyltransferase PCAF is required for Hedgehog-Gli-dependent transcription and cancer cell proliferation. Malatesta M, Steinhauer C, Mohammad F, Pandey DP, Squarrito M, Helin K. (2013) Cancer Res. 2013 Aug 13. [Epub ahead of print]

Collaborations:

The Core Facility for HT Cell Based Screens works in close collaboration with the High Throughput Translational Hematology (HTTH) Laboratory under the lead of Krister Wennerberg and Cornelia Steinhauer. This enables additional access to state-of-the-art automation technology and data analysis tools.

THE FUTURE

The facility has an open application to expand in terms of screening content, imaging capacity (confocal microscopy), plate handling and bioinformatic data analysis.

Core Facility for HT Cell Based Screens
 Biotech Research and Innovation Center (BRIC), Faculty of Health and Medical Sciences,
 Copenhagen University
 Ole Maaløes Vej 5, 2200 Copenhagen, Denmark

